



## Two novel tricyclic diterpenoids from *Isodon rubescens* var. *taihangensis*

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**Abstract**—Two novel tricyclic diterpenoids rubescensins U (**1**) and V (**2**) were isolated from the leaves of *Isodon rubescens* var. *taihangensis*. They were elucidated as a 8,15-*seco-ent*-kauranoid and an *ent*-abietanoid, respectively, by 1D and 2D NMR spectra, and single crystal X-ray analysis. Compound **1** is the first example of an 8,15-*seco-ent*-kaurane from the plants genus *Isodon*. A discussion of their biogenesis is described.

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### 1. Introduction

In recent years, a series of tricyclic diterpenoids were reported from the genus *Isodon*, which was well-known to be abundant in tetracyclic *ent*-kaurane diterpenoids.<sup>1</sup> Among them, adenanthin L (**3**) from *I. adenantha*,<sup>2</sup> laxiflorin O (**4**) from *I. eriocalyx* var. *laxiflora*,<sup>3</sup> and eriocaside A (**5**) from *I. eriocalyx*,<sup>4</sup> were elucidated as *ent*-abietanoids; Melissoidesin L (**6**) from *I. Melissoides*,<sup>5</sup> was an abietanoid, and taibaihenryiin C (**7**) from *I. henryi* was even regarded as having a novel skeleton,<sup>6</sup> on the basis of their tricyclic skeleton. In our continuing research for more bioactive substances from the *Isodon* plants, two tricyclic diterpenoids (**1** and **2**) were isolated from *Isodon rubescens* var. *taihangensis* Z. Y. Gao and Y. R. Li,<sup>7</sup> a famous folk herbal medicine for treatment of cancers.<sup>8</sup> Compounds **1** and **2** were determined as a 8,15-*seco-ent*-kauranoid and an *ent*-abietanoid by the key H-8 $\beta$  of **1** and H-8 $\alpha$  of **2**, respectively. From the similarity in the structures of compounds **1** and **2**, a brief discussion of their biogenesis is described.

### 2. Results and discussion

Compound **1** was obtained as colorless, prismatic crystals

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with a molecular formula C<sub>20</sub>H<sub>28</sub>O<sub>6</sub> determined by the HREIMS. The 20 carbon atoms found in the <sup>13</sup>C and DEPT NMR spectra of **1** consisted of a ketonic carbon, an aldehydic carbon, an olefinic quaternary carbon, an olefinic methylene carbon, a hemiacetal carbon, seven methine carbons including three oxygenated ones, four methylene carbons, two quaternary carbons, and two methyl carbons, which obviously suggested a diterpene skeleton. Compound **1** was further deduced to be a tricyclic diterpenoid by the absence of a quaternary carbon found in other typical *ent*-kauranoids also isolated from the same plant, such as lasiodonin (**8**),<sup>9</sup> and the presence of H-8 clearly exhibiting HMBC correlations with C-10, C-11, and C-13 (Table 1). Because H-8 has been determined to be of a  $\beta$  orientation by the ROESY correlations of H-8/H-5 $\beta$  and H-8/H-9 $\beta$ , and considering the structures of diterpenoids isolated from this plant, compound **1** was deduced to be a 8,15-*seco-ent*-kauranoid, instead of an *ent*-abietanoid.

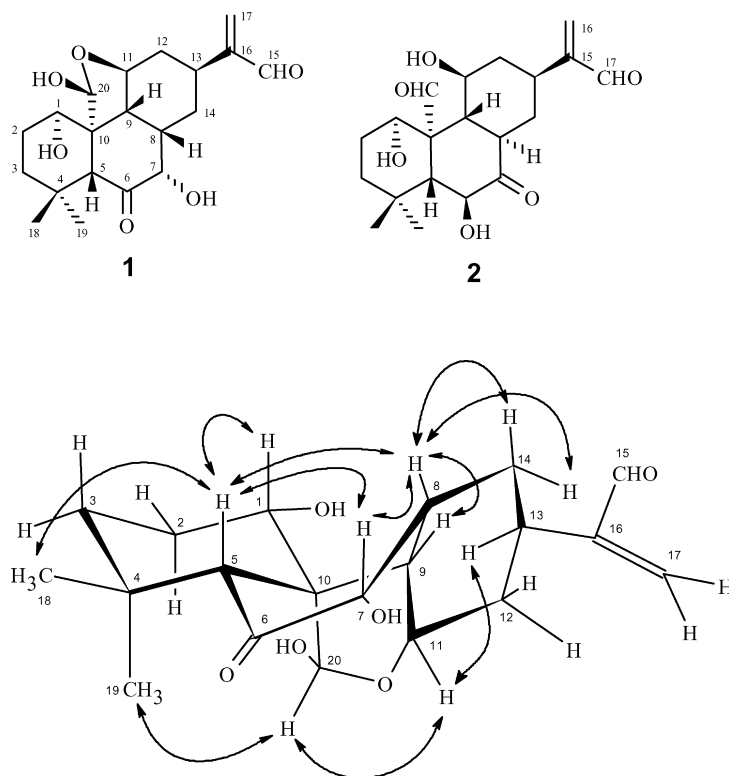
The remaining oxygenated functionalities of **1** were established accordingly. OH-1 $\alpha$  and OH-7 $\alpha$  were deduced by the HMBC correlations of H-1/C-5 and C-9, H-7/C-5 and C-9 (Table 1), and the ROESY correlations of H-1 $\beta$ /H-5 $\beta$  and H-7 $\beta$ /H-8 $\beta$  (Fig. 1). The ketonic carbon was assigned as C-6 by the long-range correlations of H-5 and H-7 with C-6 in the HMBC spectrum. Based on the analysis of the relational HMBC correlations of **1** (Table 1), the olefinic bond conjugated with the aldehydic group was located at C-13. The 11,20-epoxy group was also deduced in the same way. Consequently, with the aid of the NOEs of H-11 $\alpha$ /H-13 $\alpha$  and H-20/Me-19 in the ROESY spectrum, compound **1**

**Table 1.** NMR spectral data and HMBC correlations for **1** and **2**<sup>a</sup>

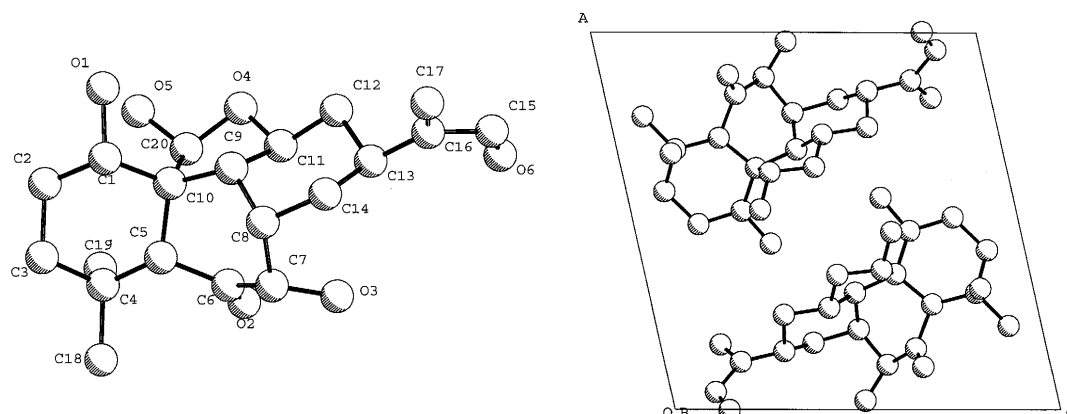
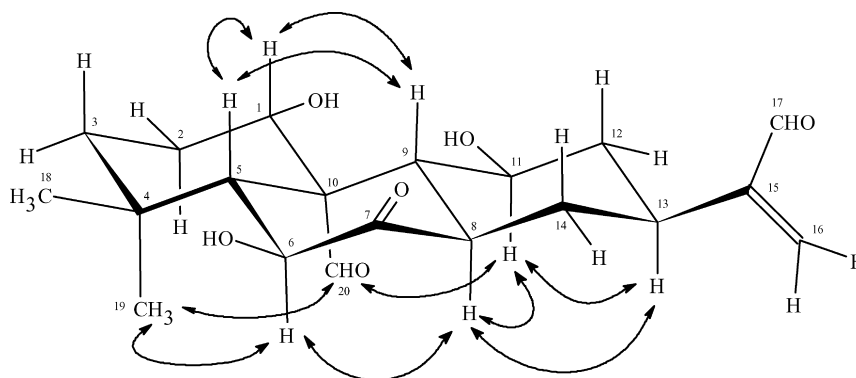
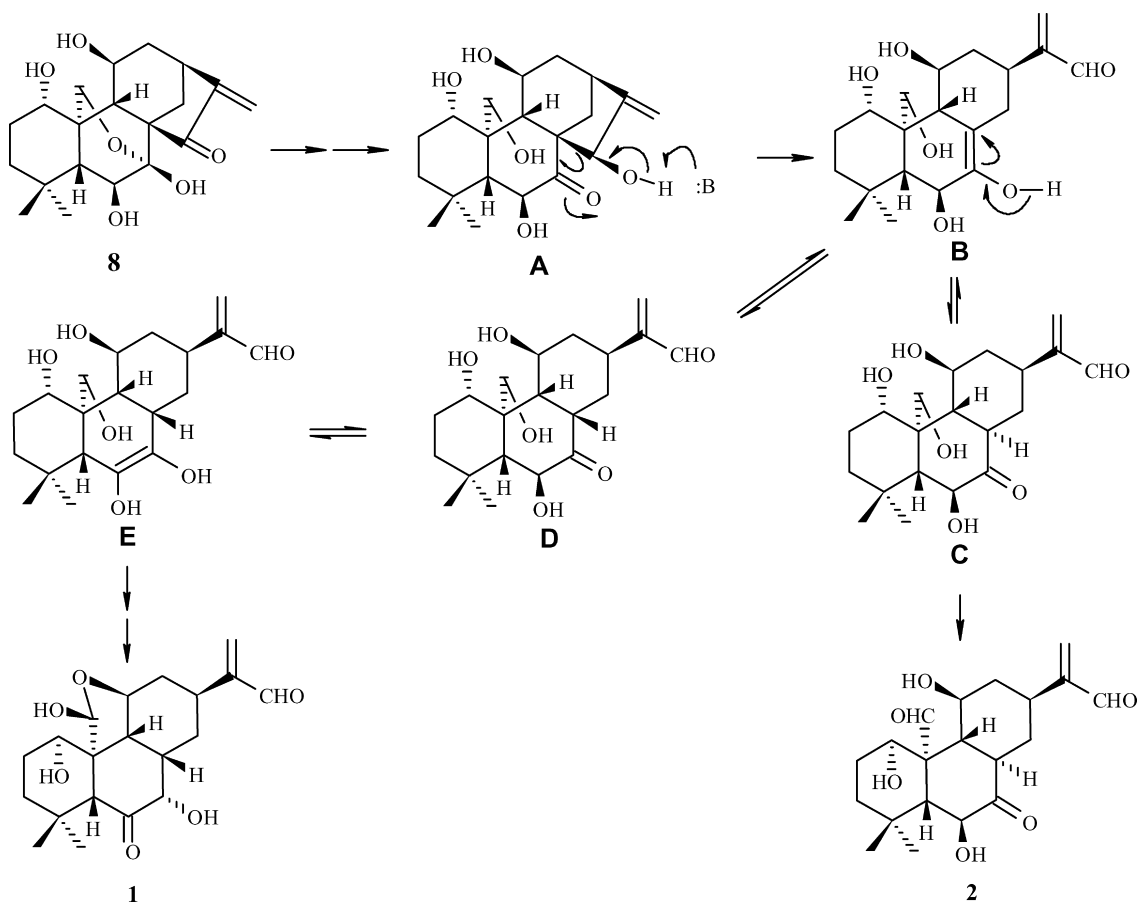
No.	<b>1</b>			<b>2</b>		
	<sup>1</sup> H	<sup>13</sup> C	HMBC <sup>b</sup>	<sup>1</sup> H	<sup>13</sup> C	HMBC <sup>b</sup>
1	3.92–3.95 m	76.9 d	9, 20	4.22–4.27 m	74.7 d	2, 3, 5, 9, 20
1-OH	6.15 d, 8.0		1, 10	7.95 s		
2	2.80–2.84 m	29.8 t	4, 10	2.10–2.15 m	28.9 t	1, 3, 4, 10
	2.00–2.06 m			2.03–2.08 m		
3	1.51 overlap	41.4 t	1, 4, 5, 18, 19	1.60–1.65 m	39.7 t	1, 2, 4, 5
	1.38 dt, 4.0, 13.2			1.39–1.45 m		
4		32.7 s			34.9 s	
5	2.49 s	57.8 d	6, 7, 9, 18, 19, 20	1.66 d, 10.0	57.5 d	6, 7, 9, 10, 18, 19
6		214.8 s		4.67 d, 10.0	75.2 d	4, 5, 7
6-OH				8.02 s		
7	4.52 d, 8.0	77.7 d	5, 6, 8, 9		210.8 s	
7-OH	6.88 s		6, 8			
8	3.08–3.11 m	37.2 s	10, 11, 13	3.37 dt, 2.2, 12.0	47.5 d	7, 9, 11, 14
9	2.68 dd, 3.6, 11.0	52.6 d	1, 5, 7, 8, 12	1.71 br t, 12.0	59.9 d	5, 8, 10, 11, 20
10		54.4 s			58.7 s	
11	3.66 dt, 3.0, 11.0	72.5 d	8, 10, 20	4.40–1.45 m	70.3 d	8, 9, 12
11-OH				5.79 br s		
12	2.32 overlap	36.7 t	9, 13, 14, 16	2.36–2.41 m	41.9 t	9, 11, 13, 14, 16
	1.58–1.63 m			1.51–1.55 m		
13	3.81–3.85 m	33.4 d	8, 11, 14, 15, 17	2.72 br t, 12.5	32.8 d	16
14	2.28 overlap	31.9 t	8, 9, 13, 16	2.25 dd, 2.2, 13.2	31.8 t	7, 8, 9, 12, 16
	1.41–1.46 m			1.45–1.49 m		
15	9.53 s	194.5 d	13, 17		153.6 s	
16		154.8 s		6.19, 5.89 (each 1H, s)	133.4 t	13, 15, 17
17	6.12, 5.84 (each 1H, s) s	133.6 t	13, 15	9.55 s	194.4 d	13, 16
18	1.01 s (3H)	30.3 q	3, 4, 5, 19	1.40 s (3H)	34.0 q	3, 4, 5, 19
19	1.67 s (3H)	21.0 q	3, 4, 5, 18	1.27 s (3H)	23.1 q	3, 4, 5, 18
20	5.73 d, 8.0	103.2 d	1, 5, 9, 11	10.73 s	207.9 d	10
20-OH	8.47 d, 8.0		10, 20			

<sup>a</sup> <sup>1</sup>H NMR, 400 MHz; <sup>13</sup>C NMR, 100 MHz, pyridine-*d*<sub>5</sub>; data in ppm (*J* in Hz).

<sup>b</sup> From H to C.



**Figure 1.** Selected ROESY correlations for **1**.

Figure 2. Crystal structure of **1**.Figure 3. Selected ROESY correlations for **2**.Figure 4. A plausible biogenetic pathway to account for the formation of compounds **1** and **2**.

was elucidated as 20(*S*)-1 $\alpha$ ,7 $\alpha$ ,20-trihydroxy-6,15-dioxo-11 $\beta$ ,20-epoxy-8,15-*seco-ent*-kaur-16(17)-ene, named rubescensin U (**1**). Finally, the X-ray crystallographic analysis of **1** (Fig. 2) confirmed stereochemically that rings A and C were in chair forms, and ring B showed a twist boat conformation. This 8,15-*seco-ent*-kaurene skeleton was proved and reported from the genus *Isodon* plants for the first time.<sup>10</sup>

Similarly, compound **2** was also deduced to be a tricyclic diterpenoid. A comparison of the <sup>1</sup>H and <sup>13</sup>C NMR data of compounds **2** and **1** suggested that **2** was derived from the hydrolysis of the hemiacetal group at C-20 of **1**. Further analysis of the ROESY spectrum of **2** revealed a key difference between **2** and **1**, in that the H-8 of **2** showed NOEs with H-13 $\alpha$  and H-11 $\alpha$  (Fig. 3) instead of correlating with H-5 $\beta$  and H-9 $\beta$ , indicating the  $\alpha$ -orientation of H-8. This presence of H-8 $\alpha$  was confirmed by the coupling constant ( $J=12.0$  Hz) between H-8 and H-9 $\beta$ , and indicated that **2** is an *ent*-abietanoid. Accordingly, by the ROESY correlation of H-6 $\alpha$ /Me-19, compound **2** was established as 1 $\alpha$ ,6 $\beta$ ,11 $\beta$ -trihydroxy-7,17,20-trioxo-*ent*-abiata-15(16)-ene, and named rubescensin V.

The biogenesis from lasiodonin **8**, one of the major *ent*-kauranoids of this plant, to compounds **1** and **2** was postulated (Fig. 4) to explain their origins. In the proposed biogenetic pathway, a retroaldol reaction from **A** to **B** resulted in the key transformation that converted a tetracyclic *ent*-kaurane to a tricyclic diterpenoid.<sup>11,12</sup> The keto-end equilibration of **B** gave **C** and **D**, and determined the key stereochemical difference between **1** and **2**. The subsequent enolization from **D** to **E**, oxidation and hemiacetalization yielded **1**. Compound **2** was derived from the oxidation of **C**. Thus, the *ent*-abietanoid **2** could have originated from an *ent*-kaurane.

### 3. Experimental

#### 3.1. General procedures

Melting points were measured on an XRC-1 micro melting point apparatus and were uncorrected. Optical rotations were measured on a JASCO DIP-370 digital polarimeter. IR spectra were obtained on a Bio-Rad FtS-135 spectrophotometer with KBr pellets. MS were recorded on a VG Auto Spec-3000 spectrometer. 1D- and 2D NMR spectra were obtained on the Bruker AM-400 and DRX-500 instruments with TMS as an internal standard.

#### 3.2. Plant material

The leaves of *Isodon rubescens* var. *taihangensis* were collected from Hebi Prefecture, Henan Province, in August 2000, and identified by Professor Z. W. Lin, Kunming Institute of Botany. A voucher specimen has been deposited in the Herbarium of the Kunming Institute of Botany, Chinese Academy of Sciences.

#### 3.3. Extraction and isolation

The 70% Me<sub>2</sub>CO extracts of the air-dried and powdered

leaves of *I. rubescens* var. *taihangensis* (10 kg) were partitioned with EtOAc to afford the EtOAc extract (400 g), which was subjected to silica gel column chromatography using CHCl<sub>3</sub>, CHCl<sub>3</sub>–Me<sub>2</sub>CO (9:1, 8:2, 7:3, 6:4) and Me<sub>2</sub>CO as eluents. Compounds **1** and **2** (14 and 6 mg) were obtained from the CHCl<sub>3</sub>–Me<sub>2</sub>CO (7:3) fraction after repeated silica gel column chromatographic separations, followed by preparative TLC and recrystallization from MeOH.

**3.3.1. Compound 1.** Colorless prismatic crystals. Mp 202–204 °C;  $[\alpha]_D^{21.6}=-60.0$  ( $c=0.1$ , acetone); IR (KBr)  $\nu_{\max}$ : 3433, 2928, 1716, 1683, 1683, 1124 cm<sup>-1</sup>. <sup>1</sup>H NMR (C<sub>5</sub>D<sub>5</sub>N, 400 MHz) and <sup>13</sup>C NMR (C<sub>5</sub>D<sub>5</sub>N, 100 MHz): see Table 1; EI-MS (70 eV)  $m/z$  (%): 364 (M<sup>+</sup>, 3), 346 (20), 328 (8), 318 (40), 300 (15); HREIMS  $m/z$ : [M]<sup>+</sup> 364.1897 (calcd for C<sub>20</sub>H<sub>28</sub>O<sub>6</sub> 364.1886).

**Crystal data for 1.** Crystals of **1**, crystallized from methanol, belong to the monoclinic space group P2<sub>1</sub>. Crystal data: C<sub>20</sub>H<sub>28</sub>O<sub>6</sub>·H<sub>2</sub>O,  $M=364.43$ ,  $a=12.368(2)$ ,  $b=6.275(1)$ ,  $c=12.289(2)$  Å,  $\beta=102.76(1)^\circ$ ,  $V=930.2(3)$  Å<sup>3</sup>,  $Z=2$ ,  $d=1.301$  g/cm<sup>-3</sup>, Mo K $\alpha$  radiation, linear absorption coefficient  $\mu=1.0$  cm<sup>-1</sup>. A colorless quadrate lumpish crystal of dimensions 0.02×0.15×0.60 mm<sup>3</sup> was used for X-ray measurements on a MAC DIP-2030 diffractometer with a graphite monochromator, maximum 2 $\theta$  value of 50.0° was set. The total number of independent reflections measured was 1530, 1431 of which were considered to be observed ( $|F|^2 \geq 8\sigma|F|^2$ ). The structure was solved by the direct method SHELX-86 and expanded using difference Fourier techniques, refined by the program and method NOMCSDP<sup>13</sup> and full-matrix least-squares calculations. Hydrogen atoms were fixed at calculated positions. The final indices were  $R_f=0.071$ ,  $R_w=0.070$  ( $w=1/\sigma|F|^2$ ).

**3.3.2. Compound 2.** White amorphous powder;  $[\alpha]_D^{21.4}=-5.0$  ( $c=0.2$ , acetone); IR (KBr)  $\nu_{\max}$ : 3441, 2928, 1705, 1683, 1084 cm<sup>-1</sup>. <sup>1</sup>H NMR (C<sub>5</sub>D<sub>5</sub>N, 400 MHz) and <sup>13</sup>C NMR (C<sub>5</sub>D<sub>5</sub>N, 100 MHz): see Table 1; (+) FAB-MS  $m/z$ : 365 ([M+1]<sup>+</sup>); (+) HRFABMS  $m/z$ : [M+H]<sup>+</sup> 365.1987 (calcd for C<sub>20</sub>H<sub>29</sub>O<sub>6</sub> 364.1964).

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